



## **The Prevalence of Gastrointestinal Parasites of Guinea Fowl (*Numida Meleagris Galeata*) Slaughtered in Kaduna Metropolis, Kaduna State, Nigeria**

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### **ABSTRACT**

*The study was conducted to determine the prevalence of gastrointestinal parasites of guinea fowl (*Numida meleagris galeata*) slaughtered in Kaduna metropolis, Kaduna State, Nigeria. A total of 200 adult Guinea fowls were sampled. The entire gastrointestinal tracts were collected. While simple flotation technique was used to collect helminth eggs in the faeces for identification. The endoparasites recovered from the gastrointestinal tracts were properly examined. Of the 200 guinea fowls examined, a total of 168 (84 %) were infected with helminth parasites. A total of 9 helminths were recovered from the infected Guinea fowls – 4 nematodes and 5 cestodes. The nematodes were *Ascaridia numidae*, *Subulura brumpti*, *Heterakis numidaenarum* and *Syngamus trachea* with prevalence of 57 % (114), 24 % (48), 10 % (20) and 10 % (20), respectively. *Raillietina tetragona*, *R. echinobothrida*, *R. cesticillus*, *R. magninumida* and *Hymenolepis carioca* were the cestodes identified with prevalence of 58 % (116), 33 % (66), 2.5 % (5), 0.5 % (1) and 0.5 % (1) respectively. 140 (70 %) of the guinea fowls shed helminth eggs in their faeces. The helminth eggs and oocysts recovered belongs to *Ascaridia*, *Capillaria*, *Syngamus*, *Heterakis*, *Raillietina*, *Strongyle* and *Eimeria* with a percentage prevalence of 55 % (110), 36 % (72), 29 % (58), 8.5 % (17), 7 % (14) and 0.5 % (1), respectively. Majority of the Guinea fowls slaughtered within Kaduna metropolis had high prevalence of gastrointestinal (cestodes and nematodes) which is a very serious problem to the guinea fowl population. It may be due to the absence of good management practices and hygienic conditions which when properly harnessed can control this menace in the Guinea fowl population in Kaduna metropolis.*

**Keywords:** Prevalence, Gastrointestinal, Tracts, Parasite, Helminths, Guinea Fowl.

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## INTRODUCTION

About 20 billion poultry exist worldwide, and of this, 75% are in developing countries, [1]. The World Bank has estimated that it will be necessary to increase meat production by about 80 percent between 2000 and 2030 [2]. Poultry production is an important means of providing high-quality protein for human consumption; as it is the second most widely consumed meat product in the world [3]. Poultry production thus represents a significant portion of the economy as a source of income for small and large holder farmers. They are not only important in narrowing the gap between the demand and supply of the proteins of animal origin but also provide an efficient means of income generation in small scale. The poultry industry was now recognized as an important sub sector of Agriculture [4-6]. When agro-economical issues and the demographics of the human population are considered, village poultry often ranks highly in terms of being an existing resource whose productivity can be increased with only a modest input [7].

In most African countries, backyard poultry accounts for more than 60% of the Total National Flock (TNF), with an asset value of more than 5.75 billion US Dollars [8]. The Guinea-fowl (*Numida meleagris galeata*, Pallas), is a common indigeneous bird on the African continent. In Nigeria, it has a population of 54.7 million and ranks second in the poultry sector, comparing favourably with domestic chicken for meat and egg production [9]. In the Northern Savannah areas of the country, it is found both in the wild and backyards where it is domesticated for meat and eggs by peasant farmers [10, 11, and 12] and as a source of income [13].

In sub-Saharan Africa, there are several species of poultry [14, 6] mainly represented by

domestic indigenous chicken (*Gallus gallus domesticus*), guinea fowl (*Numida meleagris*), duck (*Cairina sp.*) and turkey (*Meleagris gallopavo*); their distribution varies from one region to the other depending on both the physical and social environment [14].

In the last decade, the poultry industry especially chicken production has experienced a dramatic expansion. These expansions have been accompanied by the introduction of new management techniques and a large increase in the population of birds [15]. However, a high percentage of birds are local village flocks and compose mainly chicken, guinea fowls, turkeys and ducks. These flocks are managed on free range and their sources of water are mainly stagnant waters like dug out pools, ponds and wells [15].

Guinea fowl thrives well under semi-intensive conditions, forages well, and requires little attention. It retains many of its wild ancestors' characteristics: it grows, reproduces, and yields well in both cold and hot conditions. Compared to chickens, guinea fowls are economically more suitable to tropical regions because of their adaptation to traditional breeding [16]. The guinea fowl potential to increase meat and egg production among low-income countries should therefore be given greater attention [17]. However, several factors, especially those of disease, particularly of parasitic origin limit production, resulting in losses due to mortality and morbidity [18, 13, 19].

The vulnerability of Guinea fowl to parasitic infection could be attributed to its feeding on a wide range of diets due to poor husbandry, and abundance of beetles, cockroaches and earthworms that could serve as intermediate hosts, predisposing them to parasitic infections [19,20].

Multiple or mixed helminth infection has been reported as being most common in poultry kept extensively [8, 21]. Helminthes are known to compete with the infected hosts, lowering their productivity and in severe cases, causing death by blocking the gastrointestinal tract (G.I.T) especially when the worm burden is high [22].

Parasites and infectious diseases occur in nearly all ecosystems [23] and can have detrimental effects on the health status of their host [24] yet the abundance and distribution of parasites have received little attention despite their pervasiveness, diversity, and potential impacts on host population and community dynamics [25, 26, 27]. Parasitic infections (nematodes, cestodes and trematodes) often contribute to low productivity and impair the health status of poultry [28]. Parasitism is one of the major problem which inflict heavy economic losses to the poultry in the form of retarded growth, reduced weight gain, decrease egg production, diarrhoea, obstruction of intestine, poor feathers, replacement birds that take long to reach maturity, morbidity and mortality. Stress from parasites could affect the blood picture and cause anorexia [29, 30]. Investigations

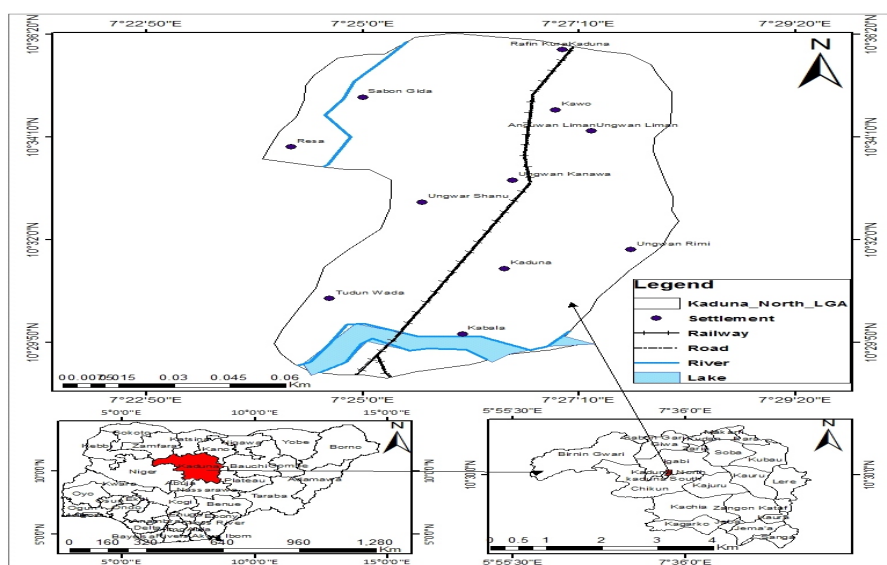
elsewhere have demonstrated the presence of some gastrointestinal and haemoparasites in poultry. [31], [32] and [11] reported *Ascaridia galli*, *Eimeria* spp. and *Heterakis gallinarum* in guinea fowls around the Kainji game park, Kwara State, Nigeria. [33] reported *Gongylonema ingluvicola*, *Raillietina* species and *Ascaridia numidae* in guinea fowls in Zaria [11], reported *Ascaridia galli* *Eimeria* spp. And *Heterakis gallinarum* in guinea fowls around the Kainji game park, Niger State, Nigeria, and [13] reported a prevalence of 45% for cestodes infection in guinea fowls in Maiduguri. Other authors have similarly made report on the prevalence of helminth parasites in guinea fowls in Nigeria [33]. According to [33], nematodes and cestodes constitute the most important group of helminth parasites of poultry in the area studied. A considerable number of arthropods, earthworms and snails, have been implicated as intermediate hosts of helminths [22]. There is dearth of information about G.I.T. parasites affecting guinea fowls in Kaduna because apart from the study carried out by [33] in Zaria, there has been no known other study.

## MATERIALS AND METHOD

### Study Area

The study was carried out in Kaduna North L.G.A, Kaduna State.

Kaduna north local Government is located at Latitude  $10^{\circ} 34' 4.80''$  N and Longitude  $7^{\circ} 27' 7.20''$  E.



Source: Ministry of Land and Survey. Kaduna State, 1992

### Sample Collection

The sample size of this study was 155 based on the prevalence 88.9 % of helminths of Guinea fowl in Zaria as reported by [33]. The samples were collected using the randomised sampling technique.

Birds of both sexes were included in the study. The sample was collected for a period of 10 weeks, twice a week and 10 samples were collected per visit. Immediately after slaughter, the alimentary tract of each bird was collected in a polythene bag, labelled appropriately and transported in a cooler containing ice block to the Helminthology Laboratory of the Department of Veterinary Parasitology and Entomology, A.B.U Zaria. Faecal materials from each bird was collected to check for helminth eggs.

### Identification of Endoparasites

The alimentary tract from the oesophagus to the cloaca was examined externally for gross lesions. The different sections of the G.I.T. were separated in Petri dishes and each section was opened longitudinally and the contents carefully washed through a sieve, helminths seen were picked out for identification. The mucosa was scraped to collect the helminthes (if present) embedded in the mucosal layer. Finally, the contents were examined under a stereomicroscope for presence and identification of helminths.

### McMaster faecal egg count method

Two gram ( 2g) of faeces was weighed and thoroughly mixed into a polythene tube. Five ml of water was added to the faeces and broken with the aid of a glass rod. The polythene tube was filled up with water, closed with the lid and shaken very well. The sample was centrifuged

at 2000 rpm for 3 minutes and the supernatant was decanted. 6 ml of the flotation medium was added, shaken thoroughly to loosen the faeces and then centrifuged at 2000 rpm for 3 minutes. The supernatant was saved in a graduated polythene tube, this was repeated twice.

After centrifuging twice, the fluid volume of the supernatant was added up to 12 ml by adding more flotation medium. Well mixed aliquot of the supernatant of each sample was charged in both chambers of the McMaster slide and FEC was done at x10 objectives. The number of helminths eggs per gram of faeces was obtained by multiplying the total number of eggs counted in the two squares of the McMaster slide by the dilution factor of 20.

### Data Analyses

Descriptive statistics was performed for all the data obtained. One-way Analysis of Variance (ANOVA) with Tukey's multiple comparison tests was performed using GraphPad Prism Version 4.00 for Windows, GraphPad Software, San Diego California USA (www.graphpad.com), values of  $p < 0.05$  were considered significant.

## RESULTS

### Prevalence of Helminth infection / Egg Count

Of the 200 Guinea fowls examined, a total of 168 (84 %) were infected with helminth parasites. 140 (70 %) of the Guinea fowls were found to shed helminth eggs in their faeces (Table 1). A total of nine (9) helminths were recovered from the infected Guinea fowls – four (4) nematodes and five (5) cestodes. The nematodes identified were *Ascaridia numidae*,

*Subulura brumpti*, *Heterakis numidaenarum* and *Syngamus trachea* with prevalence of 57 % (114), 24 % (48), 10 % (20) and 10 % (20), respectively. *Raillietina tetragona*, *R. echinobothrida*, *R. cesticillus*, *R. magninumida* and *Hymenolepis carioca* were the cestodes identified with prevalence of 58 % (116), 33 % (66), 2.5 % (5), 0.5 % (1) and 0.5 % (1), respectively (Table 2).

The helminth eggs and oocysts recovered from

the faecal samples were *Ascaridia*, *Capillaria*, *Syngamus*, *Heterakis*, *Raillietina*, *Strongyle* and *Eimeria* with a percentage prevalence of 55 % (110), 36 % (72), 29 % (58), 8.5 % (17), 7 % (14) and 0.5 % (1), respectively (Table 3). Except for 19.5 % (39) of the guinea fowls infested with a single helminth, most of the guinea fowls had mixed infestations with either two (28.5 %), three (25.5 %), four (10 %) or five (0.5 %) helminth parasites (Table 4).

**Table 1: Prevalence of parasites of Guinea fowls slaughtered in Kaduna metropolis**

	Egg count	Helminths	Non-infected
N	140	168	7
Prevalence (%)	70	85	3.5

N = number of guinea fowls infected

**Table 2: Prevalence of gastrointestinal nematodes and cestodes of guinea fowls slaughtered in Kaduna metropolis**

Parasites	Nematodes					Cestodes			
	<i>Ascaridia numidae</i>	<i>Heterakis gallinarum</i>	<i>Subulura brumpti</i>	<i>Syngamus trachea</i>	<i>R. tetragona</i>	<i>R. echinobothrida</i>	<i>R. magninumida</i>	<i>R. cesticillus</i>	<i>H. carioca</i>
N	114	20	48	20	116	66	1	5	1
Prevalence (%)	57	10	24	10	58	33	0.5	2.5	0.5
Total no. of helminths	2,804	98	639	53	3105	1,565	1	94	20
Mean per bird	24.60	4.90	13.31	2.65	26.77	23.71	1	18.80	20
Range	2 – 129	1 – 13	1 – 157	1 – 8	2 – 171	1 – 134	0 – 1	5 – 38	1 – 19

N = number of guinea fowls infected



**Table 3: Prevalence of helminth eggs and coccidia oocysts of guinea fowls slaughtered in Kaduna metropolis**

	<i>Ascaris</i>	<i>Capillaria</i>	<i>Syngamus</i>	<i>Heterakis</i>	<i>Strongyle</i>	<i>Raillietina</i>	<i>Eimeria</i> oocysts
<b>N</b>	110	72	58	17	1	14	95
<b>Prevalence (%)</b>	55	36	29	8.5	0.5	7	47.5

N = number of guinea fowls infected

**Table 4: Prevalence of guinea fowls with single or multiple infections in Kaduna metropolis**

<b>No. of species of helminths</b>	<b>N</b>	<b>Prevalence (%)</b>
<b>One</b>	39	19.5
<b>Two</b>	57	28.5
<b>Three</b>	51	25.5
<b>Four</b>	20	10.0
<b>Five</b>	1	1.0
<b>0</b>	32	16.0
<b>Total</b>	<b>200</b>	<b>100</b>

N = number of guinea fowls infected

## DISCUSSION

The overall prevalence of 84.0% recorded in guinea fowls in this study is slightly lower than those reported by [35, 36] and [21] in Guinea fowls in studies carried out in Vom Plateau, and Maiduguri, Borno States Nigeria, respectively. Some authors also reported higher prevalence rate for the local domestic chickens raised on free-range [37, 38, 39, 40, 41, 42, 43, 44, 45, 46]. On the other hand, several authors reported values that were lower than the prevalence reported in this study [47, 48, 19, 43, 49, 8, 50, and 51]. The helminth composition, prevalence and intensity reported in this study agrees with the studies of [35, 36], [37], [52], [53], [43, 54], [51] in Nigeria and [30] in Zimbabwe. This high prevalence rate could be attributed to the habit of domesticated fowls feeding on a wide

range of diets under poor husbandry, and abundance of beetles, cockroaches and earthworms that could serve as intermediate or paratenic hosts, predisposing them to parasitic infections [19, 20]. Nevertheless, some minor differences exist in the helminth composition, prevalence and intensity of individual helminths between this study and the studies of the aforementioned researchers. The minor differences in the results of the present study could be explained on the basis of seasonal, management, climatic variations and also due to variation in parasitic population of different localities where the birds were exposed [55].

The more prevalent genus of helminths was cestodes with *Raillietina tetragona* as the most prevalent. This finding is similar with the findings of [33] who reported that *Raillietina*

*tetragona* had the highest prevalence rate for guinea fowls sampled in Zaria. Similar findings [55] in Kashmir, India in a research to determine the most prevalent gastrointestinal parasites in semi-scavenging backyard poultry. The results in this study also coincides with the observations of [38] who also reported that in chicks of Amhara region of Ethiopia, among cestodes, *Raillietina tetragona* showed the highest percentage of prevalence. The percentage prevalence of *Raillietina echinobothridia* observed from this study is lower than the observations of [40] and [41] but higher than the prevalence rate reported by [44] for free-range chicken in Abeokuta. *Raillietinna cesticillus* reported prevalence rate lower that those reported by [41], [56] and [57]. In general, the most dominant cestodes were the *Raillietina* spp which had since been known several decades ago to be cosmopolitan and contributes to nutrient depletion in birds as reported by [58] and [22]. The habit of free range poultry of scratching any material including cow dung to look for ants, beetles and maggots could account for the high prevalence of *Raillietina* whose intermediate hosts including ants and beetles, are maggots of *Musca domestica* [30] which are excessively abundant in Kaduna State.

*Hymenolepis carioca* had the lowest prevalence. Similar observations (51-59) had been reported, though with *H. carioca* having a higher prevalence

The nematodes recovered from the intestines of the Guinea fowls in this study were few. This coincides with the findings of [51]. This however, was different from the findings of [46], [43], [19], [57], [60], [61], [33] and [48], who all reported higher prevalence for nematodes. Among the nematodes, *Ascaridia numidae* showed the highest percentage of

prevalence. The prevalence of *Ascaridia numidae* in this study was higher than the value reported by [33] but the values are still lower than the prevalence as reported by [35, 36], [62] and [37] in chickens. Since guinea fowls are more susceptible to the parasites, it might have accounted for the higher prevalence in this study.

*Subulura brumpti* also reported a high prevalence rate in this study. This was similar to the rate reported by [41] who found a comparable result in local domestic chickens in central Ethiopia, and [57] in North-Western Algeria.

The prevalence of *Heterakis numidaenarum* was found to be relatively similar to the prevalence rate reported by [37] and [33] in poultry and guinea fowls in Zaria, Nigeria. On the other hand, the prevalence rate in this study was lower than those reported by [45] and [41]. [57], reported *H. numidaenarum* as having the highest prevalence rate in their study conducted in Southern Nigeria, Central Ethiopia, North-western Algeria, respectively. The differences in observed could be attributed to differences in geographical location. Despite the relatively low prevalence rate of *H. numidaenarum* in this study, it poses significant danger to the avian population due to its association with the protozoan *Histomonas meleagridis*, the causal agent of “blackhead” (enterohepatitis) of turkeys [8, 63, 64]; as such guinea fowls could act as reservoirs of infection to locally domesticated chickens in this study area.

*Syngamus trachea* recorded a low prevalence rate. This is in agreement with the reports of [19] and [33] in Zaria, and [65] in Jos, who reported in their work that this parasite has low prevalence rate of infection compared to the other helminth parasites. [36] and [37] on the

other hand did not observe *S. trachea* in their study. This could be attributed to the fact that their study was conducted in a season when the conditions were not favourable for the abundance of the intermediate hosts.

The complete absence of trematodes in this study agrees with the study of [35], [53], [37], [33], [41], [19] and [66]. These findings could be associated with seasonal variations in the availability of free water which could have limited the exposure of the guinea fowls to snails, which are the intermediate host of trematodes, hence the absence in this study. Also, guinea fowls would prefer tree tops and hilly areas, these habitats could limit them from coming into contact with the gastropod intermediate host [67].

### Conclusions and Recommendations

Majority of guinea fowls slaughtered within the Kaduna metropolis have high prevalence rate of gastrointestinal (cestodes and nematodes) which pose very serious problems to the guinea fowl population. It may be due to the absence of good management practices and hygienic conditions.

The gastrointestinal parasites recovered were four nematodes (*Ascaridia numidae*, *Heterakis numidaenarum*, *Subulura brumpti*, *Syngamus trachea*) and five cestodes (*Raillietina tetragona*, *R. echinobothrida*, *R. magninumida*, *R. cesticillus* and *Hymenolepis carioca*).

The most common cestode specie is *Raillietina tetragona* with prevalence of 58 % (116) while the most common nematode specie was *Ascaridia numidae* with prevalence of 57 % (114).

This study highlights the fact that gastrointestinal parasites present a serious

challenge to the well being of guinea fowl in Kaduna.

It is suggested that appropriate parasite monitoring need to be invented in order to minimize the risks of strong infestations in the guinea fowl population in Kaduna metropolis.

Institution of a programmed control measure for improved harnessing of the potentials of guinea fowl production in Kaduna metropolis.

Poultry farmers should be encouraged to seek veterinary services in other to boost the guinea fowl production sector of agriculture.

### REFERENCE

1. FAO, Food and Agricultural Organization, (2007). Statistical database. Rome. Series No. 38.
2. FAO, Food and Agricultural Organization, (2011). Draft guidelines on phenotypic characterization of Animal Genetic resources. *Commission on Genetic resources for food and Agriculture*, 13th Regular Session, 18-22 July, 2011, Rome. Available at <http://www.fao.org/docrep/meeting/002/om651e.pdf>
3. Soniya, E. B. (1996). The context and prospects for development of small holder rural poultry production in Africa, proceeding CTA international seminar on small holder rural poultry production. *Thessaloniki Greece*, 135 – 152.
4. Pandey, V. S, Demey, F. And Verhustlst, A. (1992). *Parasitic diseases; a neglected problem in Village poultry in sub-Sahara Africa*. In: Pendy, V. S and Demey, F. (Eds). *Village poultry production in Africa* Rabat, Morocco, pp. 136 – 141.
5. Magothe, T. M., Okeno, T. O., Muhuyi, W. B. And Kahi, A. K. (2012). Indigenous chicken production in Kenya: 1. Current status.



- World's Poultry Science Journal*, 68: 119-132.
6. Yakubu, A., Abimiku, H. K., Musa-Azara, I. S., Idahor, K. O. and Akinsola, O. M. (2013). Assessment of flock structure, preference in selection and traits of economic importance for domestic turkey (*Meleagris gallopavo*) genetic resources in Nasarawa State, Nigeria. *Livestock Research for Rural Development* 25, Article #018. Retrieved, from <http://www.lrrd.org/lrrd25/1/yaku25018.htm>
  7. Copland, J. W. and Alders, R. G. (2009). The comparative advantages of village or smallholder poultry in rural development. In: *Village chickens, poverty alleviation and the sustainable control of Newcastle disease* (Alders, R. G., Spradbrow, P. B. and Young, M. P. (eds). Proceedings of an international conference held in Dar es Salaam, Tanzania, 5–7 October 2005. Published by the Australian Centre for International Agricultural Research (ACIAR), GPO Box 1571, Canberra ACT 2601, Australia. Pp. 11-14.
  8. Nnadi, P. S. And George, S. O. (2010). Cross sectional survey on parasites of chickens in selected villages in the sub humid zones of south-eastern Nigeria. *Journal of Parasitology Research*, 6.
  9. Nwagu, B. I., Fulayi, B. A. And Nwagu, F. O. (1997). Hatchability of Guinea fowl eggs in Nigeria. *Tropical Animal health production*, 63–64.
  10. Akinwunmi, A. (1981). The future of poultry in Nigeria. *World Poultry Science*, 31:1.
  11. Akinboade, O. A., Ogunji, F. O., Okaeme, A. N., Ayeni, J. S. O. and Dipeolu, O. O. (1983). *Comparative studies on the parasites of wild guinea fowls Numida meleagris pallas and domestic fowls*. In: Selected Papers from State of Knowledge Workshop on the Grey Breasted Helmet Guinea Fowl. Organized and hosted by Kainji Lake Research Institute, New Bussa. Co-Sponsored by the Federal Livestock Department and Federal Department of Forestry, Lagos.
  12. Okaeme, A. N. (1989). Gastro intestinal parasites of domestic fowl. *Journal of Animal Production, Research and Wildlife*, pp. 533-59.
  13. Biu, A. A. and Etukwudo, J. (2004). Cestodes of the guinea fowl *Numida meleagris gelata* in Borno State Nigeria. *Nigerian Journal of Experimental and Applied Biology*, 5: 2.
  14. Yakubu, A., Peters, S. O., Ilori, B. M., Imumorin, I. G., Adeleke, M. A., Takeet, M. I., Ozoje, M. O., Ikeobi, C. O. N. and Adebambo, O. A. (2012). Multifactorial discriminant analysis of morphological and heat tolerant traits in indigenous, exotic and crossbred turkeys in Nigeria. *Animal Genetic Resources*, 50: 21-27.
  15. Adekeye, J. O. (1986); Prevalence of *campylobacter fetus subspecies jejuni* in duck faeces in Zaria and Samaru towns. *Zariya Veterinarian*, 1(1): 37-39
  16. Dahouda, M., Toleba, S. S., Youssao, A. K. I., Kogui, S. B., Aboubakari, S. Y. and Hornick, J. L. (2007). Guinea fowl rearing constraints and flock composition under traditional management in Borgou Department, Benin. *Family Poultry*, 17: 3–17.
  17. Madzimure, J. Saina, H. And Ngorora, G. P. K. (2011). Market potential for guinea fowl (*Numidia meleagris*) products. *Tropical Animal Health Production*, 43: 1509–1515.
  18. Ikeme, M. M. (1970). Helminths infestation of poultry in Nigeria. *Tropical Veterinarian*, 5: 97–102.
  19. Luka, S. A. And Ndams, I. S. (2007). Gastrointestinal parasites of domestic chicken *Gallus gallus domesticus* Linnaeus 1758 in Samaru, Zaria, Nigeria. *Science World Journal*, 2(1): 27–29.
  20. Onyirioha, J. N. N. (2011). Gastrointestinal helminthes fauna of native domestic fowl (*Gallus gallus domesticus*) in Owerri area of Nigeria. *Researcher*, 3(1): 124–126.

21. Biu, A. A., Rabo J. S., Dawurung, J. S. and Lagu, A. A. (2012). Prevalence of Nematodes of Domesticated Guinea Fowl in Maiduguri, Nigeria. *New York Science Journal*, 5(3): 6–8.
22. Soulsby, E. J. L. (1982). Helminths, Arthropods and Protozoa of Domesticated Animals (Seventh Edition). *Society and Bailliere Tindall. London*, pp. 378–777.
23. Robert, L. S. And Jonovy, J. (1996). *Foundation of Parasitology*. In: Dubuque, I A. Brown Publisher.
24. Clayton, D. and Moore, J. (1997). Host-parasites evolution; general principal and avian models. Oxford, UK, Oxford University Press.
25. Windsor, D. A. (1998). Most of the species on earth are parasites. *International Journal for Parasitology*, 28: 1939–1941.
26. Poulin, R., (1999). The functional importance of parasites in animal communities: Many roles at many levels? *International Journal for Parasitology*, 29: 903–914.
27. Mouritsen, K. N., and Poulin, R. (2005). Parasites boosts biodiversity and changes animal community structure by trait-mediated indirect effects. *Oikos*, 108: 344–350.
28. Bahadory, S. R., Rad, N. H., Ramezani, A., Babazadeh, D., Falah, S. and Ghavami, S. (2014). Evaluation of Gastrointestinal Helminths of Native Turkeys in Amol, Iran. *The Journal of World's Poultry Research*, 4(4): 86-88.
29. Shah, A., Anwar A., Khan, M., Iqbal, Z. And Qudoos, A. (1999). Comparative studies on the prevalence of cestode parasites in indigenous and exotic layers at Faisalabad. Department of Veterinary Parasitology, University of Agriculture, Faisalabad, Pakistan. *International of Agriculture and Biology*, 1(4): 277-279
30. Dube, S., Zindi, P., Mbangwa, J. and Dube, C. (2010). A Study of Scavenging Poultry Gastrointestinal and Ecto-parasites in Rural Areas of Matebelel and Province, Zimbabwe. Department of Applied Biology and Biochemistry, National University of Science and Technology, Bulawayo. *International Journal of Poultry Sciences*, 9(9): 911-915.
31. Sol, D. Jovani, R. And Torres, J. (2000). Geographical variations in blood parasites in feral pigeons the role of vector, *Ecography*, 23: 307–314.
32. Adriano, A. E., and Cordeiro, S. N. (2001). Prevalence and Intensity of *Haemoproteus columbae* In three Species of Wild Doves from Brazil. *Mem. Inst. Oswaldo, Cruz, Rio de Janeiro*, 96(2): 175–178.
33. Nfor, M. B., Ajanusi, O. J., Agbede, R. I. S. And Esievo, K. A. N. (1999). Prevalence of parasites of Guinea fowl (*Numida meleagris galeata*) in Zaria – Nigeria. *Bulletin of Animal Health Production for Africa*, 47: 103 – 106.
34. Thrustfield, M. (1995). *Veterinary Epidemiology* (Second Edition), Oxford Blackwell Science London U.K., pp. 479.
35. Fabiyi, J. P. (1972a). Incidence of the helminth parasites of the domestic fowl in the Vom area of Benue-Plateau State, Nigeria. *Bulletin of Epizootic Diseases of Africa*, 20: 229–234.
36. Fabiyi, J. P. (1972b). Studies on parasites of the grey-breasted helmet guinea fowl (*Numida meleagris galeata* Pallas) of the Vom area of the Benue Plateau State, Nigeria. *Bulletin of Epizootic Diseases of Africa*, 20: 235–238.
37. Fatihu, M. Y., Ogbogu, V. C., Njoku, C. O. and Saror, D. J. (1991). Comparative study of gastrointestinal helminths of poultry in Zaria, Nigeria. *Revue d' Elevage et de Medecine Veterinaire des pays Tropicaux*, 2: 175–177.
38. Eshetu, Y. Mulualem, E., Ibrahim, H., Berhanu, A. and Aberra, K. (2001). Study of gastrointestinal helminthes of scavenging chickens in four rural districts of Amhara region, Ethiopia. *Revue Scientifique et Technique*, 20: 791–796.

39. Hove, T., Permin, A., Esmann, J. B., Hoj, C. H., Mukaratirwa, S. (2002). Ecto, endo and haemoparasites in free range chickens in the Goromonzi District in Zimbabwe. *Preventive Veterinary Medicine*, 54(3): 213 – 224.
40. Sam-Wobo, S. O. And Mafiana, C. F. (2003). Prevalence and Identification of Helminth Parasites of local chickens of Abeokuta, Nigeria. *Asset Series*, 2(2): 141 – 147.
41. Ashenafi H. and Eshetu Y. (2004). Study on gastrointestinal helminths of local chickens in Central Ethiopia. *Revue Médecine Vétérinaire*, 155: 504–507.
42. Yoriyo, K. P., Fabiyi, J. P., Panda and Adamu, S. U. (2005). Intensities of helminth parasites of free ranging chickens in Bauci and environs. *Yankari Journal*, 2: 135–139.
43. Yoriyo, K. P., Adang, K. L., Fabiyi, J. P and Adamu, S. U. (2008a). Helminthes parasites of local chickens in BauchinState, Nigeria. *Science World Journal*, 3: 35–37.
44. Eslami, A., Ghaemi, P. and Rahbari, S. (2009). Parasitic infections of free-range chickens from Golestan Province, Iran. *Iran Journal of Parasitology*, 4: 10–14.
45. Ekpo, U. F., Ogbooye, A. A., Oluwole, A. S. and Takeet, M. (2010). A preliminary survey on the parasites of free range chicken in Abeokuta, Ogun State, Nigeria. *Journal of Natural Sciences, Engineering and Technology*, 9(2): 123–130.
46. Mwale, M. And Masika, J. P. (2011). Point prevalence study of gastrointestinal parasites in village chickens of Centane district, South Africa. *African Journal of Agricultural Research*, 6: 2033–2038.
47. Azare, B. A. (1997). Prevalence of helminth parasites of domestic chickens (*Gallus domesticus*). M.Sc. Thesis, Abubakar Tafawa Balewa University, Bauchi, Nigeria.
48. Matur, B. M. (2002). Prevalence of some gastrointestinal parasites in pullets of chickens (*Gallus gallus domestica*) in the Federal Capital Territory Abuja. *Journal of Tropical Bioscience*, 2: 78–82.
49. Tolossa, Y. H., Shafi, Z. D. and Basu (2009). Ectoparasites and gastrointestinal helminthes of chickens of three agro-climatic zones in Oromia Region, Ethiopia. *Journal of Animal Biology*, 59: 289–297.
50. Dawet, A., Yakubu, D. P., Daburum, Y. H., Dung, J. P. and Haledu, U. I. (2012). Gastrointestinal helminthes of domestic chickens (*Gallus gallus*) in Jos, Plateau State, Nigeria. *Nigerian Journal of Parasitology*, 33: 85–89.
51. Adang, K. L., Asher, R. and Abba, R. (2014). Gastro-intestinal helminthes of domestic chickens (*Gallus gallus domestica*) and ducks (*Anas platyrhynchos*) slaughtered at Gombe main market, Gombe State, Nigeria. *Asian Journal of Poultry Science*, 8(2): 32–40.
52. Elizabeth, N., Gadzama, E. N. and Srivastava, G. C. (1986). Prevalence of intestinal parasites of market chickens in Borno State. *Zariya Veterinarian*, 1(2): 126–128.
53. Gadzama, E. N. And Strivastrava, G. C. (1986). Prevalence of gastrointestinal parasites of market chickens in Borno State, Nigeria. *Zariya Veterinary*, 1: 126–128.
54. Yoriyo, K. P., Adang, K. L., Adamu, S. U. and Sambo, Z. (2008b). Prevalence of gastrointestinal helminthes in free-ranginh chickens and guinea fowls in Bauchi and its environs. *Bulletin of Pure and Applied Science*, 27: 1–6.
55. Salam, S. T. (2015). Gastro-intestinal helminthiasis: an unseen threat to the backyard poultry production of Kashmir valley. *International Journal of Science and Nature*, 6(1): 63–69.
56. Hassouni, T. And Belghyti, D. (2006). Distribution of gastrointestinal helminths in chicken farms in the Gharb region, Morocco.

*Parasitology Research*, 99: 181–183.

57. Yousfi, F., Kheira, S., Ilyes, M., Hanene, D. and Touria, H. S. (2013). Gastrointestinal helminths in the local chicken *Gallus gallus domesticus* (Linnaeus, 1758) in traditional breeding of North-Western Algeria. *Biodiversity Journal*, 4(1): 229–234.

58. Cheng, T. (1973). *General Parasitology*. Academic Press, New York, San-Francisco, pp. 434–936.

59. Ilyes, M. And Ahmed, B (2013). Cestode parasites of free-range chickens (*Gallus gallus domesticus*) in the North-Eastern of Algeria. *International Journal of poultry science*, 12(11): 681–684

60. Kose, M., Sevimli, F. K., Kozan, E. K. And Cicek, H. S. (2009). Prevalence of gastrointestinal helminthes in chicken in Afyonkarahisar district, Tuckey. *Kafkas University Vet. Fakultesi Dergisi*, 15: 411–416.

61. Rayyan, A., Al-Hindi, A. And Al-Zain, B. (2010). Occurrence of gastrointestinal helminthes in commercial and free-range chickens in Gaza strip, Palestine. *Egypt. Poultry Science*, 30: 601–606.

62. Okon, E. D. And Enyenihi, N. U. (1980). A study of parasite of local fowls in Oron, Cross River State, Nigeria. *Nigeria Journal of Parasitology*, 1(2): 82–86.

63. Calnek, B. W., Barnes, H. J., Bread, C. W., Reid, W. M. and Yoder, J. H. W. (1991). *Diseases of Poultry*, (Ninth edition). Iowa State, University Press Ames, Iowa, U.S.A., pp. 745–751.

64. Tibor, K. I. (1999). *Veterinary Helminthology*. Butterworth, Heinemann Oxford (First Edition). pp. 138–141, 188-184.

65. Pam, V. A., Daniel, L.N., Brengshak, S., Wai, M. S., Omalu, C. J. And Ashi, R. D. (2006). The survey of intestinal parasites of local and exotic chickens slaughtered at Yankari market, Jos, Plateau State. *Journal of Medical and Pharmaceutical Sciences*, 2(3): 27.

66. Mahairwa, A. P., Msoffe, P. L., Ramadhani, S., Mollel, E. L., Mtambo, M. M. A. And Kassuku, A. A. (2007). Gastrointestinal helminthes in free-range ducks in Morogoro Municipality, Tanzania. *Livestock Research and Rural Development*, 19: 4.

67. Puttalakshamma, G. C., Ananda, K. J., Prathiush, P. R. Mamatha, G. S. And Suguna, R. (2008). Prevalence of Gastrointestinal parasites of poultry in and around Bangalore. *Veterinary World*, 1(7): 201–202.